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# The unified approach of radiation on biological matter: Action of sparsely ionizing

# radiation on mammalian cells

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### Abstract

Using a wide range of published data from irradiation experiments by sparsely ionizing radiation over mammalian cells, a tangible relationship between the cellular damage; effect cross-section and radiation quality represented the spacing between ionizing events over the path of generated primary electrons; the mean free path for linear primary ionization. The maximum damage by sparsely ionizing radiation can only be established by ultra-soft x-rays from C (k) where the mean free path along the track of the cell nucleus is about 2 nm. At higher mean free path, other ultra-soft and soft x-rays become potentially harmful only with the contribution of water radicals. A simple semi-theoretical model is proposed to define absolute biological effectiveness based on effect inactivation cross section which is interrelated to the mean free path for linear primary ionization. For sparsely ionizing radiation, the model shows a feasible saturation region for the effect cross-section 5  $\mu$ m<sup>2</sup> for mean free path less than 1.8 nm. The model explains the mechanisms leading to cell death via DNA strand scissions. The effect of sparsely ionizing radiation contributes only to a fraction of (1/20) of the maximum reachable damage.

Keywords: Effect cross-sections; Mean free path; Mammalian cells; Sparsely ionizing radiation; Biophysical modelling

### 1. Introduction

Although heavy charged particles (termed as densely ionizing radiation; DIR) have its own advantages over conventional x-ray, y-ray and electron beams (termed as sparsely ionizing radiation; SIR) in the therapeutic treatments, especially for deep situated tumors, SIR are still important in other areas of medical sciences including radiography, mammography computer tomography and even therapeutic treatments [1]. Depending on their energy, photons propagate through matter and interact with its atomic electrons. For energies up to 100 keV, the photon is completely absorbed by the atomic electron via photo-electric effect. At energies up to few MeV, Compton scattering becomes the predominant interaction, where the photon gives up part of its energy to the electron. With higher energies up, pairproduction will be more important. In all processes, primary electrons are produced. The fate of these free electrons inside biological matter is decided by their kinetic energy. They lose energy through ionizations and excitations events while slowing down in biological matter. The average energy lost per ionization along the electron track in water is within the range of 22 - 26 eV [2]. The average distance between successive ionization is longer, for higher energy electrons and deceases as the electron slows down. As whole, primary electrons produce sparse ionization tracks as compared to dense tracks produced by heavy charged particles like protons or other heavier charged ions. Different radiation qualities such as track Linear Energy Transfer (LET) and the mean free path for linear primary ionization (MFP;  $\lambda$ ) can be characterized by the ionisation density. LET is defined as the mean energy deposited per track length and is measured in (keV/µm), while MFP represents the mean spacing distance between two consecutive ionizations along the primary track measured in (nm) [3-4]. Unlike DIR which are characterized by their high LET values, SIR identified by their lower LET values. LET of x-rays with values 0.1 – 20 keV/µm depend on x-rays spectrum, which is

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determined by a combination of three factors, the x-ray tube anode material, the x-ray beam filtration material and the high operational voltage (KV). LET for  $\gamma$ -rays is about 0.5 – 1. KeV/ $\mu$ m [5 - 6]. LET of high energy photons produced in linear accelerators varies between 0.15 – 4.5 keV/ $\mu$ m for electron energies varies between 50 – 4 MeV. These LET values are substantially low as compared with heavy charged particles LET, i.e. for 4 MeV  $\alpha$ -particles, LET is about 100 KeV/ $\mu$ m.

Deoxyribonucleic acid (DNA) is widely accepted as the vital target for the induction of cell killing by ionizing radiation. Upon their interaction with DNA molecules, ionizing radiations produce a variety of damages, including double-strand breaks (dsb's), single-strand breaks (ssb's) and base damages, as presented in Figure-1 a. Due to their high ionization density, DIR interact directly with DNA molecules to induce adjacent DNA breaks forming what is known as dsb's. On the other hand, Photons and energetic electrons interact with DNA molecules either directly with lower probability, or indirectly via ionization of water molecules with higher probability (as compared to DIR) to generate a spectrum of lesions, of which base damage and ssb's are the most abundant [7]. The enhancing role of water radicals produced by SIR interactions with water to damage the DNA is shown in Figure-1b. In the last few decades, researchers have provided evidences that the double breaks of DNA opposite strands are responsible for cell death [8-11].



**Figure 1** (a) Schematic representations of selected types of ionizing radiation damage to the DNA; ssb, dsb (two adjacent ssb's) and base damage. (b) Direct and indirect damage by SIR to the DNA where free radicals ( $OH \oplus$ ,  $H \oplus$  or hydrated electron formed by the water hydrolysis; the radicals then diffuse to damage the DNA by chemical reaction

The correlation between biological effects and a suitable physical parameter led to propositions of many biophysical models [12-14]. While the main objective of each model is to predict the shortage of experimental outcomes, it is also providing a deeper insight of the damage mechanisms at cellular level [15-17]. Watt and his group suggested that the spacing of ionizing events along primary charged particle tracks can explain the details of the different mechanisms at nanometric scales in terms of MFP for linear primary ionization [18].

Cell death is commonly considered as a reference endpoint to characterize the action of ionizing radiation in different subcellular targets. It is also the goal for any tumor therapy. With this respect, radiobiological experiments involving *in vitro* exposure of mammalian cells to different types of SIR, still provide a useful source of information. The damaging effect of what is known as reproductive death as an end point presented by survival curves; where fraction of survival usually on logarithmic scale, against radiation dose in linear scale. The shape of the curve depends on radiation type and to a certain extent on Eukaryotic cells type. In general SIR such as x-rays, and fast electrons have shoulder type of curve, while heavy charged particles such as protons, alpha particles have linear response type. It is useful to make use of linear quadratic fitting formula;

$$F(S) = \ln(S/So) = -\alpha D - \beta D^2,$$

where  $\alpha$  (Gy<sup>-1</sup>),  $\beta$  (Gy<sup>-2</sup>) are constant parameters for any specific curve. The first term represent the slope of survival curve at zero dose, which is also known as the radio sensitivity parameter. For linear survival curve;  $\beta$ = 0, and the equations becomes simply as F(S) = ln(S/So) = - $\alpha$ D.

The present study will focus on physical parameterization of the biological damage caused by different ionizing radiations. Hence presenting a model that unifies the action of SIR on mammalian cells.

#### 2. Method of Approach

It is far much better to quantify the cellular damage induced by ionizing radiation in terms of the probability of producing damage in units of area, inactivation cross section;  $\sigma_{s}(\mu m^2)$  Inactivation cross sections  $\sigma_{s}$  (in  $\mu m^2$ ) of a variety of mammalian cells were calculated using the relation:

$$\sigma_s = \frac{L_T}{6.25\rho D_o}$$

Where  $L_T$  is the track average LET (in keV/µm) for the equilibrium spectrum of charged particles involved,  $D_o$  (in Gy), and P (in gm/cm<sup>3</sup>) is the density of biological matter. Cross sections were determined for the initial slope of survival curves to avoid any problems associated with cell recovery. The initial slope for cell survival curve is simply the slope of curve at zero dose. For both survival curves whether shouldered or linear types, the initial slope is equivalent to  $\alpha$ . Hence the inactivation cross section is evaluated at  $D_o = 1/\alpha$ . For wet cells, the density of medium is assumed of water.

The cell survival radio-sensitivity parameter  $\alpha$  (Gy<sup>-1</sup>) for mammalian cell lines by the various SIR were extracted from published data [19-41]. The present study include mammalian inactivation data caused by SIR of different qualities; ultra-soft and (0.277 keV C<sub>k</sub>, 1.456 keV Al<sub>k</sub>, 4.51 keV Ti<sub>k</sub>, and Cr<sub>k</sub> 5.4 keV), soft x-rays (10 KVp – 40 KVp), medium hard x-rays (50 KVp - 110 KVp), hard x-rays (varied 120 kVp - 300 kVp), very hard x-rays (4 MV - 50 MV), <sup>60</sup>Co, <sup>137</sup>Cs  $\gamma$ -rays, and energetic electrons (~1 MeV and 18 keV), as tabulated in table-1 below. The track average structure parameters; Average energy of electrons emitted assumed to be emitted from a radiator source; E<sub>s</sub> (keV), the average LET (keV/µm) and  $\lambda$  (nm)), are all estimated using Watt's group foundations [42]. Here the average values are calculated for the electron equilibrium spectrum. The corresponding cross sections  $\sigma$  (µm<sup>2</sup>) were estimated using the above formula and the results are tabulated in Table-1.

No.	Source/Type	Es(keV)*	LET (keV/µm)	α(Gy-1)	λ(nm)	σs(μm²)	Reference
1	0.277 keV C(K) X-rays	0.1202	18.34	0.6220	2.45	1.8252	Goohead, 1980
2	1.486 keV Al(K) X-rays	0.4137	15.010	0.5233	3.23	1.2568	Goohead, 1980
3	4.510 keV Ti(K) X-rays	2.1050	8.534	0.4532	6.66	0.6188	Goohead, 1980
4	5.4 keV Cr(K) X-rays	2.6200	7.645	0.3364	7.55	0.4115	Hiebera, 1990
5	10 KVp X-rays	5.5000	3.500	0.4600	11.01	0.2576	Lehnert, 2008
6	25 KVp X-rays	7.8000	5.800	0.2260	9.02	0.2097	Lehnert, 2006
7	29 KVp X-rays	8.01	6.343	0.0760	8.57	0.0771	Heyes, 2004
8	40 kVp X-rays	8.01	7.495	0.0780	7.54	0.09354	Hoshi, 1988
9	50 kVp X-rays	7.77	8.113	0.2580	6.52	0.3346	Todd, 1965
10	55 kVp X-rays	7.59	8.193	0.2900	6.46	0.3802	Spadinger, 1992
11	80 kVp X-rays	6.79	8.347	0.2700	6.39	0.3606	Napolitanok, 1992
12	100 kVp X-rays	6.69	8.102	0.2390	6.62	0.3098	Kroneberg, 1991
13	150 kVp X-rays	7.65	6.980	0.1900	7.80	0.2122	Chang, 1992
14	180 kVp X-rays	8.98	6.310	0.1370	8.80	0.1383	Tolkendorf, 1983
15	200 kVp X-rays	10.26	5.823	0.1280	9.61	0.1193	Belli, 1989
16	210 kVp X-rays	11.42	5.590	0.1358	10.10	0.1215	Hall, 1972
17	225 kVp X-rays	12.65	5.280	0.1015	10.80	0.0858	Rodrriguez, 1981
18	250 kVp X-rays	14.68	4.772	0.1475	11.95	0.1126	Chapman, 1971

**Table 1** SIR track structure data;  $E_s$  (keV), LET (keV/mm), and  $\lambda$  (nm) along with radio-sensitivity parameter  $\alpha$  (Gy<sup>-1</sup>);  $E_s$  is the average energy of electrons produced by photons (or electron) interactions

19	300 kVp X-rays	17.81	4.421	0.1280	12.96	0.0905	Nias, 1969
20	4 MV X-rays	40.00	2.200	0.1220	29.00	0.0429	Zackrisson, 1989
21	6 MV x-ray	55.00	1.700	0.1100	37.00	0.0299	Chithrani, 2010
22	50 MV x-ray	500.00	0.420	0.148	180.00	0.0099	Zackkrisson, 1989
23	Cs-137 γ-rays	150.70	0.985	0.2474	65.52	0.0298	Hei, 1988
24	Co-60 γ-rays	335.60	0.607	0.1400	111.10	0.0099	Perris, 1986
25	0.94 MeV Y-90 β-rays	508.20	0.531	0.0780	157.40	0.0066	Barendsen, 1962
26	4 MeV electrons	1947.00	0.287	0.1282	276.00	0.0059	Haskins, 2020
27	11 MeV electrons	5058.00	0.283	0.2100	292.50	0.0077	Spadinger, 1992
28	18 MeV electrons	7876.00	0.284	0.0553	297.50	0.0025	Haskins, 2020

The search for a model imply the multi-stage trials of a function F= F ( $\lambda$ ), provided that the semi-empirical formula fits the trend observed by the  $\sigma$  ( $\lambda$ ) in terms of the justifiable physical parameters  $\lambda$ 



## 3. Results and discussion

Figure 2 The effective cross section  $\sigma$  ( $\mu$ m<sup>2</sup>) for mammalian cells vs. mean free path for primary ionization  $\lambda$  (nm) for SIR (S; soft, US; ultra-soft, MH; medium hard, VH; very hard)

Effective cross sections;  $\sigma_s$  (µm<sup>2</sup>) for the various mammalian cells including human, hamster cells as a function of  $\lambda$ (nm) for SIR, as depicted in the Table-1, shown in Figure-2 in log-log scale. The meddling of inactivation cross-sections data as well as the radiation quality parameters, as seen in the Table-1 and Figure-2, is healthy. The behaviour of each class for each type of SIR data is shown in the figure. Beside that all experimental radiobiological data through the years were performed at different laboratories, the interfering is mostly due to the nature of x-rays spectrum which are produced by different anodes, filters and HV's, radiation quality parameters for different type of SIR may be interfere. High energy electrons with HV x-ray from linear accelerators are also inclusive.

Visual inspection of log-log scale, clarifies the grouping of  $\sigma$  and  $\lambda$  data within the spread of the physical and biological errors. Based on earlier studies by the author [43], an inflection point is expected at  $\lambda_0$ =1.8 nm, attributed to the mean chord of the strands in the DNA segment and identify that the dsb's of the DNA as the critical lesion to inactivate cells for all ionizing radiation types.

The overall shape of the relation between  $\sigma$  and  $\lambda$  support the idea of having unifying response of mammalian cells to SIR with an inflection point at  $\lambda_0 = 1.8$  nm which is equivalent to the interspacing distance between DNA strands. Only C<sub>k</sub> ultra-soft x-rays with photons of energy around 0.278 keV can induce highest biological damage near the inflection point where  $\lambda_0 = 1.8$  nm. Soft x-rays with maximum operating voltage around 40 KVp shown next to US x-rays, with ionization separated by 8 nm and lower effective cross-sections. Medium and hard x- rays and  $\lambda$ -rays, as expected have much lower cross sections due to their high energy photons and their lower ionization density. SIR like hard x-rays,  $\gamma$ -rays and electrons have much smaller cross sections, as shown in Figure-2.



Figure 3 The unified model of radiation action on mammalian cells by SIR, manifested by  $\sigma$  (µm2) vs.  $\lambda$  (nm) as indicated by the dashed and solid curves presented

In searching of a mathematical model to manipulate this data, the following simple semi-empirical relation is used:

$$\sigma(\lambda) = \sigma_0 (1 - \exp(\lambda / \lambda_0)^n)$$

Where  $\sigma_0$  is the maximum reachable saturation cross section if ever exist, and  $\lambda_0$  is the value of the mean free path at inflection point (the spacing between the DNA strands). The result is presented by the solid line in Figure-3.

The gradient of this linear portion in the log-log scale is -1.59  $\pm$  0.06. This part of the curve is attributed to the repairable ssb's of the DNA. The maximum damage if ever established by ultra-soft x-rays, along with water radicals' enhancement, will be around  $\sigma_s = 5 \pm 0.6 \ \mu m^2$ 

The merit of both values; the projected saturation cross section  $\sigma_0 = 5 \ \mu m^2$  and the mean free path  $\lambda_0 = 1.8 \ nm$  (with n = 1.6) indicate that the nature of this damage has got to be related to nanometric events. The scales of these events indicate that the induction of dsb's near the inflection point i.e. C (k) US x-rays is plausible. Whereas nearby soft x-rays contribute to the induction ssb's of the DNA. The crucial rule of generated water radicals in close proximity to soft x-rays can induce dsb's of DNA. This means soft x-rays, medium and hard x-rays,  $\gamma$ -rays or even fast electrons will never reach saturation region by their own, and the dominant interactions produced are ssb's of the DNA, in the lower region of the linear part.

#### 4. Conclusion

In this work a simple model is presented. The main features of this model relies on its specification of the cellular damage in terms of biophysical parameters that relates molecular events such dsb's of the DNA to macroscopic biological effects such as cell death. In simple mathematical form;  $\sigma(\lambda) = \sigma_0 [1 - \exp(-\lambda_0/\lambda_0)^n]$ , the model indicate that the maximum damage is represented by  $\sigma_0$  (5 µm<sup>2</sup>). In theory, the highest saturation damage leading to the death of mammalian cells should be equivalent to the geometrical cross section,  $\sigma_g$  (60 µm<sup>2</sup>). SIR can only be attributed to 1/12 of the geometrical cross-section.

Further investigations are needed to indicate the rules of water radicals in cellular damage. A more sophisticated model will be vital to further demonstrate the action of ionizing radiation on mammalian cells in terms of additional radiochemical parameters related to the damaging effects by water radicals.

#### References

- [1] Khan FM, JP Gibbons, Khan's the Physics of Radiation Therapy. 5<sup>th</sup> ed. Philadelphia; Lippincott Williams & Wilkins. 2014.
- [2] Ding elder M, RH Ritchie, JE Turner, W Friedland, HG Paretzke, Hamm RN. Comparisons of Calculations with PARTRAC and NOREC: Transport of Electrons in Liquid Water. Radiat Res. 2008; 169(5): 584-594.
- [3] ICRU Report 16, Linear Energy Transfer. Inter. Comm. on Radiat. Units and Meas. 7910, Woodmont Ave. Bethesda, MD. 20814, USA, 1970.
- [4] Watt DE. A Unified System of Radiation Bio-Effectiveness and its Consequences in Practical Application. Radiation Protection Dosimetry. 1997; 70(1-4): 529–536.
- [5] Hall EJ, AJ Giaccia. Radiobiology for the Radiologist. 8th ed. Philadelphia; Lippincott Williams & Wilkins. 2018.
- [6] Kellerer AM. Electron spectra and the RBE of x-rays. Radiat Res. 2002; 158: 13-22.
- [7] Ward JF. DNA damage produced by ionizing radiation in mammalian cells: identities, mechanisms of formation, and reparability. Prog Nucleic Acid Res Mol Biol. 1988; 35: 95-125.
- [8] Ward JF. The yield of DNA double-strand breaks produced intracellularly by ionizing radiation. Int J Radiat Biol. 1990; 57(06): 1141-1150.
- [9] Iliakis G. The role of DNA double strand breaks in ionizing radiation-induced killing of Eukaryotic cells. Bioessays. 1991; 13(12): 641-648.
- [10] Pouget J-P, SJ Mather. General aspects of the cellular response to low- and high-LET radiation. Euro J Nucl Med & Molec Imag. 2004; 28(4): 541-561.
- [11] Ward JF. The complexity of DNA damage- relevance to biological consequences. Int J Radiat Biol. 1994; 66: 427-432.

- [12] Katz R, B Ackerson, M Homayoonfar, SC Sharma. Inactivation of cells by heavy ion bombardment." Radiat Res. 1971; 47: 402-425.
- [13] Kiefer J. On the interpretation of heavy ion survival data. In: Booz J & HG Ebert, eds. Proceeding of the 8<sup>th</sup> Symp. On Microdosimetry. Luxemburg: Cmm. of the Europ. Comm. 1982; 729-742.
- [14] Kampf G, Eichhorn K. DNA strand breaks by different radiation qualities and relations to cell killing: Further results after the influence of  $\alpha$ -particles and carbon ions. Studia Biophys. 1983; 93(01): 17-26.
- [15] Kramer M, G Karft. Heavy ion track structure calculations. In: Chadwick KH, Moschini G & MN Varma. Eds. Workshop on Biophysical Modelling of Radiation Effects. Bristol: Adam Hilger. 1992; 61-68.
- [16] Harder D, P Virsik-Peuckert, E Bartels. Theory of pairwise lesion interaction. In: Chadwick KH, G Moschini & MN Varma eds. Workshop on Biophysical Modelling of Radiation Effects. Bristol: Adam Hilger. 1992; 179-184.
- [17] Kellerer, AM, HH Rossi. The theory of dual radiation action. Curr. Top. Radiat. Res. Q. 1972; 8: 85-158.
- [18] Watt DE, I Al-Affan, CZ Chen, GE Thomas. Identification of Biophysical Mechanisms of Damage by Ionizing Radiation. Radiat. Prot. Dosim. 1985; 13: 285-294.
- [19] Goodhead DT, J Thacker, R Cox, RE Wilkinson. Effectiveness of 0.3 keV carbon ultrasoft x-rays for the inactivation and mutation of cultured mammalian cells. Int J Radiat Biol Relat Stud Phys Chem Med. 1979; 36(2): 101-114.
- [20] Hieber L, K Trutschler, J Smida, M Wachsmann, G Ponsel, AM Kellerer. Radiation-induced cell transformation: Transformation efficiencies of different types of ionizing radiation and molecular changes in radiation transformation and tumor cell lines. Environ Health Presp. 1990; 88: 169-174.
- [21] Lehnert A, E Lessmann, J Pawelke, W Dörr. RBE of 25 kV x-rays for the survival and induction of micronuclei in the human mammary epithelial cell line MCF-12A. Radiat Environ. Biophys. 2006; 45: 253–260.
- [22] Lehnert A, E Lessmann, J Pawelke, W Dörr. RBE of 10 kV x rays determined for the human mammary epithelial cell line MCF-12A. Radiat Res. 2008; 169(3): 330–336.
- [23] Heyes GJ, AJ Mill. The neoplastic transformation potential of mammography x-rays and atomic bomb spectrum radiation. Radiat. Res. 2004; 162(2): 120-127.
- [24] Hoshi M, S Antoku, N Nakamura, W Russell, R Miller, S Sawada, M Mizuno, S Nishio. Soft X-ray dosimetry and RBE for Survival of Chinese Hamster V79 cells Int J Radiat Biol. 1988; 54(4): 577-591.
- [25] Todd PW. Heavy-ion irradiation of cultured human cells. Radiat. Res. 1967; 7: 196-207.
- [26] Spadinger I, B Palci. The relative biological effectiveness of <sup>60</sup>Co γ-rays, 55 kVp x-rays, 250 kVp x-rays, and 11 MeV electrons at low doses. Int J Radiat Biol. 1992; 61(3): 345-353.
- [27] Napolitano M, M Durante, G Grossi, M Pugliese & G Gialanella. Inactivation of C3H 10T1/2 cells by monoenergetic high-LET α-particles. Int J Radiat Biol. 1992; 61(6): 813-820.
- [28] Kronenberg A. Perspectives on fast-neutron mutagenesis of Human Lymphoblastoid cells. Radiat Res Suppl. The international colloquium on neutron radiation biology.1991; 128(1): S87-S93.
- [29] Chang PY, CA Tobias, EA Blakely. Protein synthesis modulates the biological effectiveness of the combined action of hyperthermia and high-LET radiation. Radiat Res. 1992; 129(3): 272-280.
- [30] Tolkendorf, E, K Eichohorn. Effect of ionizing radiation of different linear energy transfer of the induction of cellular death and of chromosome aberrations in cells of the Chinese hamster. Studia Biophys. 1983; 95: 43-56.
- [31] Belli M, R Cherubini, S Finotto, G Moschini, O Sapora, G Simone, MA Tabocchini. RBE-LET relationship for the survival of V79 cells irradiated with low energy protons. Int J Radiat Biol. 1989; 55(1): 93-104.
- [32] Hall EJ, W Gross, RF Dvorak, AM Kellerer, HH Rossi. Survival curves and age response functions for Chinese hamster cells exposed to x-rays or high LET α–Particles. Radiat Res. 1972; 52(1): 88–98.
- [33] Rodriguez A, EL Alpen. Cell survival in spheroids irradiated with heavy-ion beams. Radiat Res. 1981; 85(1): 24-37.
- [34] Chapman JD, RG Webb, J Borsa. Radiosensitization of mammalian cells by p-nitroacetophenone; I. Characterization in asynchronous and synchronous populations. Int J Radiat Biol Relat Stud Phys Chem Med. 1971; 19(6): 561-573.

- [35] Nias AH, D Greene, M Fox, RL Thomas. Effect of 14 MeV monoenergetic neutrons on HeLa and P388F cells in *vitro*. Int J Radiat Biol Relat Stud Phys Chem Med. 1968; 13(5): 449-456.
- [36] Zackkrisson B, B Johansson, P Östbergh. Relative biological effectiveness and Oxygen enhancement ratio of 50 Mv X-Rays, Acta Oncologica. 1989; 28(4): 529-535.
- [37] Chithrani DB, S Jelveh, F Jalali, M van Prooijen, C Allen, RG Bristow, RP Hill, DA Jaffray. Gold nanoparticles as radiation sensitizers in cancer therapy. Radiat Res. 2010; 173: 719–728.
- [38] Hei TK, DJ Chen, DJ Brenner, EJ Hall. Mutation induction by charged particles of defined linear energy transfer. Carcinogenesis. 1988; 9: 1233-1236.
- [39] Perris A, P Pialoglou, AA Katsanos, EG Sideris. Biological effectiveness of low energy Protons. I. Survival of Chinese Hamster Cells. Int J Radiat Biol Relat Stud Phys Chem Med. 1986; 50(6): 1093-1101.
- [40] Barendsen GW. Dose-survival curves of Human cells in tissue culture irradiated with α-, β-, and 20-KV. X- and 200-KV. X-radiation, Nature. 1962; 193: 1153–1155.
- [41] Haskins JS, SK Martinez, M Engstrom, M Murakami, T Mori, D Leary, TA Kato. Electron scattering in conventional cell flask experiments and dose distribution dependency. Scientific Reports. 2020; 10(1): 482-490.
- [42] Watt DE. Quantities for Dosimetry of Ionizing Radiation in Liquid Water. London, Taylor and Francis. 1996.
- [43] Watt DE, AS Alkharam. Charged particle track structure parameters for application in Radiation Biology and Radiation Chemistry. Int. J. Quantum Chem. 1994; 52(S21): 195- 207.